Section Two
Urine Toxicology

2.4 Liquid-Liquid Extraction Methods for GC/MSD Confirmation
2.4.4 Qualitative 11-nor-9-THC-Δ^9-COOH (Carboxy-THC)

2.4.4.1 BACKGROUND

Cannabis sativa use dates back to 2700 B.C.[2,5] Marijuana (MJ) refers to a mixture of the leaves and flowering tops.[3] The smoke from burning cannabis includes 61 different cannabinoids.[2,6] The major active ingredient in marijuana is delta–9-tetrahydrocannabinol (Δ^9-THC). The Δ^9-THC content varies from 2 to 10% with an average of four to five percent. The quality of marijuana is reported to have improved over the last 20 years due to superior cultivation practices. The medicinal effects of MJ include anti-nausea, muscle relaxing, anticonvulsant and reduction of intraocular pressure.[6] Cannabis therefore has found use as an antiemetic to deal with the nausea associated with anticancer chemotherapy and for relief for those suffering from glaucoma. The debate continues on medical use and the complete legalization of the drug.

Several factors come into play when considering the behavioral effects of (Δ^9)-THC. These include the route of administration (smoked or ingested), THC concentration of the plant (dose), the experience of the user, the user's vulnerability to psychoactive effects, and the setting of the use.[5,6] The desirable effects of MJ include an increased sense of well-being, mild euphoria, relaxation and a mild sedative-hypnotic effect.[5,6] Its clinical effects are similar to those of alcohol and the antianxiety agents.[5] The side-effects of MJ use include impairment of cognitive functions, alteration of the user's perception of time and distance, reaction time, learning and short-term memory.[2,5,6] MJ has been shown to interfere with a person's ability or willingness to concentrate. Cannabis causes temporal disintegration such that the individual loses the ability to store information in the short term and is easily distracted.[2] Impairment from use is thought to last from 4 to 8-hours with more recent studies reporting 3 to 6 hours. Dr. Huestis reported that most behavioral and physiological effects return to baseline within three to six hours after use with residual effects in specific behaviors for up to 24 hours.

Impairment of coordination and tracking behavior has been reported to persist several hours beyond the perception of the high.[6] Due to the variable period of impairment, the relating of urine Carboxy-THC to the time of use, and thus impairment, requires the development of the scenario surrounding the stop for DUI. The presence of Carboxy-THC in urine only indicates exposure to MJ at some previous, indeterminate time.

The physiological effects may include an increase in heart rate and blood pressure, conjunctival suffusion, vasodilation, dry mouth and throat and a decrease in respiratory rate. The individual may also experience increased hunger (munchies).

Δ^9-THC is rapidly metabolized to the inactive metabolite, Carboxy-THC.[1,4,5,6] In urine, this major metabolite, Carboxy-THC is pursued due to Δ^9-THC only being present in minute quantities.[6] Carboxy-THC in urine has been conjugated with glucuronic acid to improve excretion. The detection time of Carboxy-THC in urine following marijuana use varies dependent upon various pharmacological factors such as the dose obtained, the route of administration, and the rates of metabolism and excretion.[1] Δ^9-THC is deposited in body fat due to its high lipid solubility. It is slowly released from this storage depot over time.[1] The amount of Δ^9-THC stored in fat is a function of the amount, frequency and potency of drug exposure. The detection time can therefore vary from days to months.

2.4.4.2 SCOPE

This method is to qualitatively confirm the presence of a major metabolite of marijuana, Carboxy-THC, in urine specimens.

2.4.4.3 EQUIPMENT AND SUPPLIES

2.4.4.3.1 Tube Rocker
2.4.4.3.2 Laboratory Centrifuge capable of 3500 rpm
2.4.4.3.3 Waterbath
2.4.4.3.4 Drybath
2.4.4.3.5 Evaporative Concentrator equipped with nitrogen tank.
2.4.4.3.5 pH Indicator Strips
2.4.4.3.6 Glassware
16X100mm tubes
16X144mm tapered tip centrifuge tubes (optional)
Snap caps for 16mm OD tubes (optional)
GC/MS ALS vials
GC/MS vial microinserts
2.4.4.3.7 Gas Chromatograph equipped with a mass selective detector and a nonpolar capillary column (e.g. 100%-dimethylpolysiloxane or 95%-dimethyl-polysiloxane with 5%diphenyl).

2.4.4.4 REAGENTS

Refer to manual section 5.12 for solution preparation instructions. Purity of chemicals must be ACS Grade or equivalent.

2.4.4.4.1 1.0 N KOH

2.4.4.4.2 Saturated Potassium Phosphate Monobasic pH ≈1.8

2.4.4.4.3 87:13 Hexane with Ethyl Acetate (v/v)

2.4.4.4.4 Ethyl acetate

2.4.4.4.5 Silylating Agent (select from)
BSTFA/1% TMCS or MSTFA

2.4.4.5 STANDARDS

2.4.4.5.1 Stock Standard Solution
100µg/mL (+) 11-nor-9-carboxy-Δ^9-THC

2.4.4.5.2 Working Standard Solution (1800ng/mL)
Add 180µL Stock Solution to 9.82mL Methanol. Other volumes may be prepared. *Solution is stable for 1-year when stored under refrigeration.*

2.4.4.6 QUALITATIVE CONTROLS

2.4.4.6.1 Positive Controls
A minimum of two spiked 60ng/mL and one commercial Carboxy-THC containing control must be analyzed in each batch of samples.

2.4.4.6.1.1 60ng/mL Carboxy-THC Spiked Controls
Add 3mL of the same lot of negative urine used to prepare the negative control to extraction tube. Add 100µL of working standard solution, and vortex.

2.4.4.6.1.2 Suitable nominal concentration range for commercial control is 15ng/mL to 150ng/mL.

2.4.4.6.2 Negative Control
Negative urine commercially obtained or in-house urine verified to be negative for drugs of interest.

2.4.4.7 PROCEDURE

2.4.4.7.1 Initial set-up
Label extraction tubes, tapered bottom derivatization tubes and GC/MS vials with microinserts for the negative control, spiked positive controls, commercial positive control(s), and casework samples.

2.4.4.7.2 Sample Preparation

Transfer 3 mL urine specimen, negative urine, spiked positive controls and commercial positive control(s) to extraction tubes.

2.4.4.7.3 Sample Hydrolysis

- Add 0.5mL 1.0N KOH to each extraction tube.
- Vortex *gently* to mix.
- Check resulting pH.
- pH must be ≥ 12. If pH<12, add additional 0.5mL of KOH.
- Place in 40°C water bath for 15 minutes.
- Allow samples to cool before proceeding with solvent extraction.

2.4.4.7.4 Extraction

If original pH was ≥ 12:

- Add 1.5mL Saturated Phosphate Buffer (pH 1.8).
- Add 3mL Hexane/Ethyl Acetate (87:13).
- Rock for 10 minutes.

If original pH was <12:

- Add 3.0mL Saturated Phosphate Buffer (pH 1.8).
- Add 4mL Hexane/Ethyl Acetate (87:13).
- Rock for 10 minutes.

2.4.4.7.5 Centrifuge tubes at ≅3500 rpm for 10 minutes.

2.4.4.7.6 Transfer upper organic phase from tube into labeled tapered bottom tube.

2.4.4.7.7 Evaporate solvent to dryness, under a gentle stream of nitrogen, at ≅37°C.

2.4.4.7.8 Derivatization

- To dried extract in tapered bottom tubes, add 50µL ethyl acetate and 50µL silylating reagent.
- Cap tubes with snap caps.
- Vortex.
- Heat tube for 15 minutes in 95°C dry bath.
- Remove from heat and allow to cool.
- Transfer derivative to labeled GC/MS ALS vial with microinsert.

2.4.4.8 GAS CHROMATOGRAPHY/MASS SPECTROMETRY (GC/MS)

2.4.4.8.1 Preparation for Analysis Run

2.4.4.8.1.1 Into Sequence log table, enter information for case samples, controls and pre-sample solvent blanks. A 60ng/mL spiked positive control must run both early and late in the sequence.

2.4.4.8.1.2 Load case samples, controls and solvent blanks into the quadrant rack(s) as noted in the sequence table.

2.4.4.8.2 GC-MSD Acquisition Parameters

2.4.4.8.2.1 Refer to instrument METHOD for current acquisition parameters.

2.4.4.8.2.2 Current acquisition method must be stored centrally as a hard or electronic copy.

2.4.4.8.2.3 Acquire sample data in SIM (selected ion monitoring) utilizing the ions 371, 473 and 488.

2.4.4.8.3 Detection and Identification Criteria

2.4.4.8.3.1 **Retention Time**

Identification requires a peak within ±0.1 minutes of the retention time established for Carboxy-THC with the in-run control(s).

2.4.4.8.3.2 **Ion ratios - Selective Ion Monitoring (SIM)**

Carboxy-THC Ion ratio for the early and late 60 ng/mL controls must be calculated and averaged. This mean ratio must be compared to ratio obtained from casework and the mean of the 60ng/mL control samples. Ratio between monitored ions, 371:473 and 371:488, must agree within ±20%.

2.4.4.8.3.2.1 Incorrect Ratios

If the casework or control sample ion ratios do not agree within ±20% due to high concentration of c-THC in the sample, the extract may be diluted with 100µL ethyl acetate. Once the extract has been diluted, control samples *and* the diluted case sample should be re-analyzed with the SIM GC/MS method. Alternatively, carboxy-THC in the sample may be confirmed using full scan

data, provided a derivatized reference material is also run in full scan mode. The analyte may be confirmed from full scan data if there are no significant differences in the mass spectral data as compared to the appropriate reference material *and* the retention time is within ±0.1 minutes of the appropriate reference material.

2.4.4.8.3.3 **Assessment of relative strength of case sample to 60 ng/mL control.** The response of case samples will be compared to a 60 ng/mL control sample. The analyst will pick one of the spiked controls and divide the response in that control by 5; this will be defined as the approximate minimum response. The approximate minimum response will be noted in the analyst's notes and a notation will be placed identifying the control that is used. The analyst will compare this response to the response for each case sample. If the response for the case sample is less than the approximate minimum response established by the control, Carboxy-THC will generally not be confirmed. If it is below the minimum response, it is at the analyst's discretion whether or not to call the drug. Other factors such as enzyme screen results and the sample response in relation to the baseline must be considered and noted in the analyst's notes.

2.4.4.9 QUALITY ASSURANCE REQUIREMENTS

2.4.4.9.1 Refer to toxicology analytical methods 5.8 and 5.10 for additional quality assurance and reference material authentication requirements.

2.4.4.10 ANALYSIS DOCUMENTATION

2.4.4.10.1 Case results are to be recorded in the LIMS system.

2.4.4.10.2 Original data for controls will be compiled for each analysis run and must be stored centrally in the laboratory where the analysis was performed, until archiving or destruction.

2.4.4.10.3 A copy of data for controls may be stored electronically in a central location and need not be included in individual case files. When necessary, a copy of control printouts can be prepared from the centrally stored document.

2.4.4.11 REFERENCES

2.4.4.11.1 Huestis, M.A., Mitchell, J.M. and Cone, E.J. *Detection Times of Marijuana Metabolites in Urine by Immunoassay and GC-MS* J. Anal. Tox. **19**:443-449, 1995.

2.4.4.11.2 Huestis, M. *Marijuana.* pp. 269-304. *in:* Principles of Forensic Toxicology, Third Edition. Levine, B. ed., AACC, 2010.

2.4.4.11.3 *Cannabis. in: Clark's* Isolation and Identification of Drugs pp. 423-425, Moffat, A.C. ed., Pharmaceutical Press:London, 1986.

2.4.4.11.4 Drug Evaluation and Classification Training Manual, U.S. Dept. of Transportation, 1993.

2.4.4.11.5 Julien, R.M. Marijuana*: A Unique Sedative-Euphoriant-Psychedelic Drug. in:* A Primer of Drug Action. pp. 319-349, W.H. Freeman and Company:NewYork, 1998.

2.4.4.11.6 O'Brien, C.P. *Drug Addiction and Drug Abuse*. pp. 572-573. *in:* Goodman & Gilman's The Pharmacological Basis of Therapeutics, Ninth edition, Hardman, J.G. ed., McGraw-Hill, 1996.

Revision History

Section Two
Urine Toxicology

2.4 Liquid-Liquid Extraction Methods for GC/MSD Confirmation
2.4.4 Qualitative 11-nor-9-THC-Δ^9-COOH (Carboxy-THC)

Revision No.	Issue Date	History
1	11-27-2001	Original Issue in SOP format
2	09-13-2002	Clarification of detection and identification criteria and refinements
3	05-07-2008	Updated QA references and language
4	07-28-2008	Clarified that negative urine used to prepare positive control is the same lot as used for negative control.
5	03-07-2011	Clarified detection and identification criteria, minor reformatting
6	12-16-2011	Removed highlighting of a previous change, changed retention time criteria from +/- 0.2 to +/- 0.1 min. Removed QC requirements covered in another method.
7	11-28-2012	Added option to dilute samples that are too strong and overload the detector.
8	2-12-13	Added minimum response criteria, clarified ratios to compare.
9	1-16-2014	Added option of confirm strong samples by full scan instead of SIM. Amendment to 2.4.4.10 in accordance with new LIMS system. Minor formatting changes
10	04/02/2015	Removed extraneous procedure summary from method scope. Clarified instrument acquisition and acceptance criteria language. Minor formatting and grammar corrections.